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¹H{¹⁵N} heteronuclear correlation and ¹⁵N cross-polarized magic angle spinning NMR studies of the coordination modes in Zn(II) chloride complexes with purine and methylpurines

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Abstract

The ¹H{¹⁵N} heteronuclear correlation and ¹⁵N cross-polarized magic angle spinning NMR spectra of purine (1) were measured. The Zn(II) chloride complexes with 1, 7-methylpurine (2) and 9-methylpurine (3) were synthesised and studied by analytical methods, far-IR and ¹⁵N NMR. The formulae of ZnL₂Cl₂ (L = 1) and [ZnLCl₂]_n (L = 2, 3) were found. The formation of Zn(II)–N coordination bonds resulted in 3–15 ppm low-frequency shifts of the respective ¹⁵N NMR signals: N-7 for 1; N-1, N-9 for 2; N-1, N-7 for 3.

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1. Introduction

Purine (1), 7-methylpurine (2) and 9-methylpurine (3) (Fig. 1) are well known heterocycles studied by NMR [1].

The data on their ¹⁵N chemical shifts (in such solvents as H₂O/D₂O, aqueous NaOH, aqueous D₂SO₄, dmso d_6 , CF₃COOH, HSO₃F) and their ²J_{NH} coupling constants were obtained by one-dimensional NMR techniques [2–5]. In this work we present the ¹H{¹⁵N} heteronuclear correlation and ¹⁵N cross-polarized magic angle spinning (CPMAS) spectra of **1**, as well as the results of a HSQC-HECADE experiment allowing for precise calculation of both ²J_{NH} and ⁴J_{NH} values.

Compounds 1-3 possess several potential N-donor metallation sites and easily form complexes with transition metals [6–13]. Their NMR studies involved mainly

¹H measurements of aqueous solutions containing 1–3 and such ions as Zn^{2+} , Cu^{2+} , Co^{2+} , Ni^{2+} , Mn^{2+} [14– 17]. The equilibrium of N(1), N(7) and N(9)-coordinated forms was generally suggested and supported by selective broadening of ¹⁵N signals in the paramagnetic $Cu^{2+} + 1$ system [18]. In the following we focus on the complexes of Zn(II). Previous one-dimensional ¹⁵N NMR measurements with the ligands mentioned above were performed for unidentified species in dmso- d_6 solutions of ZnCl₂, CdCl₂, HgCl₂ [19]. We now report the ¹H{¹⁵N} heteronuclear correlation spectra for welldefined Zn(II) chloride complexes of 1–3, dissolved in dmf- d_7 or dmso- d_6 .

Three types of Zn(II) coordination compounds, formed in aqueous or ethanol solutions of ZnCl₂ and 1, were described. Speca et al. [6] reported [Zn(purine)Cl₂]_n as the main product from the reaction of ZnCl₂ with 1 in absolute ethanol and suggested its di-, oligo- or polymeric structure with N-3, N-9 bridges. Laity and Taylor [20] obtained, by evaporation of a water solution of ZnCl₂, 1 and diluted HCl, monomeric Zn(purine)₂Cl₂

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Fig. 1. Purine (1), 7-methylpurine (2) and 9-methylpurine (3).

and proved by X-ray analysis its N-7 monodentate coordination. Sheldrick [21] reported the X-ray structure of another species, Zn(purinium)Cl₃, protonated at N-1 and Zn-metallated at N-7, this compound being isolated from a more concentrated HCl solution.

We have performed the $ZnCl_2+1$ reaction in 96% ethanol. Although the conditions were similar to those described by Speca et al. [6], a coordination compound of the formula $Zn(purine)_2Cl_2$ (1a), analogous to that described by Laity and Taylor [20] was obtained. The identity of both species was proven by us using X-ray powder diffraction. On the other hand, the Zn(II) chloride complexes with 2 and 3 revealed the formulae of Zn(7-methylpurine)Cl₂ (2a) and Zn(9-methylpurine)Cl₂ (3a).

Compounds 1a-3a were studied by us using ${}^{1}H{}^{15}N{}$ two-dimensional NMR methods (GHSQC, GHMQC), in order to determine the metallation sites and Zn(II)-induced ${}^{15}N{}$ coordination shifts. For 1a, the ${}^{15}N{}$ CPMAS and far-IR spectra in the solid phase as well as ${}^{1}H{}$ NMR, UV–Vis and conductivity measurements in aqueous solutions were also performed.

2. Experimental

2.1. Materials

The chemicals used for syntheses were $ZnCl_2$ (POCh Gliwice, Poland, 98–99%) and 1, 2,6-dihydroxy-3,7-dimethylpurine, 5-amino-4,6-dichloropyrimidine (Aldrich, 97–99%).

2.2. Methods

¹H{¹⁵N} NMR spectra were measured with a Varian INOVA 500 MHz spectrometer, equipped with a Nalorac Z-SPEC IDG 500-5HT gradient probe. The resonance frequencies were 499.805 MHz for ¹H and 50.664 MHz for ¹⁵N, the reference standards being TMS and external CH₃NO₂, respectively. The solvents were dmf- d_7 (1, 1a) and dmso- d_6 (2, 3, 2a, 3a), the concentrations being 0.1 M. The temperature was 303 K. The ¹H one-dimensional NMR spectra of aqueous solutions (D₂O) of 1 and 1a were measured with a Varian Gemini-XL 200 MHz spectrometer.

The ¹H{¹⁵N} NMR spectra of **2**, **3**, **2a** and **3a** were performed using gradient-enhanced heteronuclear multiple-quantum correlation (GHMQC) [22] with the following parameters: $\pi/2$ pulse lengths of 8.0 µs for ¹H, 24.5 µs for ¹⁵N; relaxation delay 1.5 s; long-range couplings evolution delay 50 ms (J = 10 Hz); sweep width 10 ppm in F2 and 240 ppm in F1; 1024 experiments in t1; 2K points in t2; quadrature detection in t2; linear prediction to 2048 and zero filling up to 4K real points in F1 with sine bell apodisation in both dimensions.

The ¹H{¹⁵N} NMR spectra of **1** and **1a** were detected using gradient-enhanced heteronuclear single-quantum correlation (GHSQC) [23], the conditions being analogous to those of the GHMQC measurements and completed by echo-antiecho in t1 and apodisation with $\pi/2$ -shifted squared sine bell in both dimensions.

Compounds 1 and 1a were also studied by ${}^{1}H{}^{15}N{}$ GHSQC-HECADE, a GHSQC-based two-dimensional sequence, allowing for accurate and sensitive determination of heteronuclear coupling constants $(J_{\rm NH})$ from E.COSY-type cross peaks [24,25]. Sixteen FIDs were coherently added for each echo and anti-echo data set for 256 t1/t1* increments (t1 and t1* being the evolution times of the S-spin chemical shift and heteronuclear couplings, respectively). The maximum t1 value was set to 31.6 ms, t1* incremented up to 126.4 ms, t2 sampled up to 1.02 s (in order to increase the coupling separation: shift differences ratio by a factor of 4). The optimisation was done for a heteronuclear coupling constant of 8 Hz (60 ms delay), the apodisation with $\pi/2$ shifted sine bell in t1 and 1 Hz exponential broadening in t2.

The ¹⁵N CP MAS spectra of **1** and **1a** were measured with a Bruker AVANCE-DRX 500 MHz spectrometer using a Bruker MASVTN500SB BL4 probehead and 4 mm zirconia rotors. A 10 kHz spinning speed was applied. A ramp [26] was implemented on the proton channel of the cross-polarization pulse sequence. A ¹H decoupling rf field strength of 73.5 kHz (3.4 μ s π /2 pulse length) and a two pulse phase-modulation decoupling sequence [27] were applied. The following parameters were used: frequency 50.689 MHz; spectral width 23 kHz; contact time 6 ms; acquisition time 15 ms for **1** and 25 ms for **1a**; relaxation delay 300 s for **1** and 30 s for **1a**; number of acquisitions 196 for **1** and 476 for **1a**. The spectra were referenced to solid glycine and recalculated to nitromethane (δ (NH₂ in glycine) = -347.6 ppm).

The X-ray powder diffractogram of **1a** was recorded with a Philips X-Pert Pro diffractometer, whereas that of Laity's compound, Zn(purine)₂Cl₂ [20], was simulated with a SHELX-97 computer program [28]. The far-IR spectra of **1a**-**3a** were measured in the 400–100 cm⁻¹ range with a Perkin–Elmer Spectrum 2000 FT-IR spectrometer using polyethylene discs. The UV–Vis spectra of **1** and **1a** were detected in 0.1 M aqueous solutions with a Hewlett-Packard 8453 spectrophotometer. The conductivity measurements were performed with an Elmetron CX-732 conductometer. The amount of C, H, N in 1a-3a was determined by elemental analysis, whereas that of Zn by titration with EDTA (after mineralization).

2.3. Syntheses

The synthesis of **2** from 2,6-dihydroxy-3,7-dimethylpurine and **3** from 5-amino-4,6-dichloropyrimidine followed literature procedures [19]. Compound **1a** was obtained from ZnCl₂ and **1** in 96% EtOH, whereas **2a**, **3a** from ZnCl₂ and **2**, **3** in water. The white precipitates were filtered after 24 h, washed with EtOH and dried in air. The elemental analyses revealed the formulae of Zn(purine)₂Cl₂ for **1a** (*Anal*. Calc. for C₁₀H₈Cl₂N₈Zn/ Found)—C, 31.9/32.0; H, 2.1/2.0; N, 29.8/30.2; Zn, 17.4/17.5% and Zn(methylpurine)Cl₂ for **2a**, **3a** (*Anal*. Calc. for C₆H₆Cl₂N₄Zn/Found for **2a**/Found for **3a**)— C, 26.6/26.5/26.8; H, 2.2/2.3/2.4; N, 20.7/20.5/20.5, Zn 24.2/24.0/24.0%.

3. Results and discussion

3.1. ${}^{1}H{}^{15}N{}$ NMR spectra of 1-3

The ${}^{1}H{}^{15}N{}$ GHSQC spectrum of **1** (in dmf- d_7) is presented in Fig. 2.

We observe the following ${}^{15}N{}^{-1}H$ correlations via two bonds: N-1 with H-2 and H-6; N-3 with H-2; N-7 with H-8; N-9 with H-8. Some weaker ${}^{15}N{}^{-1}H$ spinspin interactions via four bonds, N-3 with H-6 and H-8, have also been detected. These results allow for an unambiguous assignment of ${}^{15}N$ signals, identical with that known from one-dimensional NMR experiments



Fig. 2. ${}^{1}H{}^{15}N{}$ GHSQC spectrum of 1 (in dmf- d_7).

[5], their chemical shifts decreasing in order N-1 > N-3 > N-7 > N-9. It is worth noting that N-7 and N-9 correlate in the same way. They can be easily distinguished, however, as the latter nitrogen possess a higher shielding constant due to its pyrrole-type character [1].

The ¹⁵N chemical shifts of **1** (Table 1), recorded in dmf- d_7 , differ not more than 2 ppm from those found in dmso- d_6 [4,5]. The ¹⁵N–¹H coupling constants, determined from the HSQC-HECADE spectrum, are as follows: ² $J_{N1H2} = 15.3$, ² $J_{N1H6} = 10.8$, ² $J_{N3H2} = 15.2$, ² $J_{N7H8} = 10.9$, ² $J_{N9H8} = 10.0$, ⁴ $J_{N3H6} = 1.4$, ² $J_{N3H8} = 1.5$ Hz. The values of ² J_{NH} differ less than 1 Hz from the literature data [5], whereas those of ⁴ J_{NH} do not exceed a typical [29] magnitude of 1.5 Hz.

The ¹H{¹⁵N} GHMQC spectra of **2** and **3** have been performed in dmso- d_6 , the ¹⁵N chemical shifts (Table 1) being identical with those described in literature [4,5].

3.2. Spectroscopic properties of 1a-3a

3.2.1. X-ray powder diffraction and far-IR studies of 1a-3a

In spite of many attempts, we were not able to obtain monocrystals of **1a**, suitable for X-ray analysis. Therefore, we have measured the X-ray powder diffractogram of **1a** and compared it with that simulated for Laity's compound, Zn(purine)₂Cl₂, based on the published atomic coordinates [20]. The most intensive reflections, measured at 15.5°, 20.8°, 26.0°, 27.3° (2 θ angles), appear also in the simulation (Fig. 3), allowing for a conclusion that both complexes are one species.

In the far-IR spectrum of **1a** two strong absorption bands at 321 and 303 cm⁻¹ have been detected. They derive, most likely, from the Zn–Cl stretching vibrations, typical for pseudotetrahedral monomeric molecules of the ZnL₂Cl₂ type [30]. The noted v_{ZnCl} frequencies are similar to those reported for Zn(pyridine)₂Cl₂ (329, 297 cm⁻¹) [31] or Zn(imidazole)₂Cl₂ (304, 291 cm⁻¹) [32]. The far-IR spectra of **2a** and **3a** are more difficult to interpretate due to the broad absorp-

Table 1		
The ¹⁵ N chemical shifts (ppm) of 1-3 and 1a-3a (coordination	shifts
in parentheses)		

Compound	N-1	N-3	N-7	N-9
1 ^a	-101.9	-120.9	-169.3	-196.7
1a ^a	-110.7	-121.4	-179.6	-199.1
	(-8.8)	(-0.5)	(-10.3)	(-2.4)
2 ^b	-102.3	-109.4	-237.6	-137.0
2a ^b	-106.6	-110.8	-235.2	-146.8
	(-4.3)	(-1.4)	(+2.4)	(-9.8)
3 ^b	-103.4	-130.0	-140.8	-230.8
3a ^b	-106.4	-128.7	-155.8	-228.2
	(-3.0)	(+1.3)	(-15.0)	(+2.6)

^a In dmf- d_7 .

^b In dmso-*d*₆.



Fig. 3. X-ray powder diffractogram of 1a (top) and simulation for Laity's compound (bottom).

tion band in the $150-350 \text{ cm}^{-1}$ range, associated most likely with the polymeric structure of both complexes.

3.2.2. Aqueous solution of 1a

The mixture of $ZnCl_2$ and 1 in water did not yield any precipitate, contrary to the case of 2 and 3, where the insoluble complexes of 2a and 3a were rapidly formed. The reason is that 1a, which easily precipitates in ethanol, is unexpectedly well soluble in water. Its aqueous solution reveals physical and spectroscopic properties identical with those of the $ZnCl_2$ and 1 mixture.

The water solutions of either **1a** or $ZnCl_2+1$ conduct electricity with molar conductivity characteristic for Zn^{2+} and 2 Cl⁻. Their UV–Vis spectra exhibit the same absorption band at 263 nm and the molar absorbance of free **1** [33]. Their ¹H NMR spectra, measured in D₂O, are slightly dependent on the concentration (H-8 8.34, H-2 8.65, H-6 8.82 ppm for 0.1 M), being similar to that of pure **1** [34,35].

These observations suggest that monomeric **1a** easily dissociates in water into Zn^{2+} , $2 Cl^{-}$ and 2 molecules of **1**. In other words, in aqueous medium **1** has no ability to bind Zn(II) ions. The process of dissociation of **1a** is entirely reversible; however, when water is evaporated, the obtained solid residue appears to be pure **1a**, not the ZnCl₂+**1** mixture. Such a behaviour seems to be quite unusual in the case of a coordination compound.

In dmf and dmso, 1a-3a dissolve without dissociation because the respective solutions reveal the conductivity analogous to the pure solvents.

3.2.3. ${}^{1}H{}^{15}N{}$ NMR spectra of 1a-3a

The ¹H{¹⁵N} GHSQC and GHMQC spectra of 1a-3a were measured in dmf- d_7 or dmso- d_6 , 1a being also studied by HSQC-HECADE. They can be assigned in the same way as those of 1-3, the respective ¹⁵N chemical shifts being collected in Table 1 (coordination shifts in parentheses).

For 2a and 3a the shielding of two nitrogen atoms: N-1; N-9 (2a) and N-1; N-7 (3a) has been observed.

Because both complexes reveal the formula of $ZnLCl_2$ (L = 2 or 3), one can assume that the methylpurines behave as bidentate ligands and bridge the Zn(II) ions via N-1 and N-9 (2a) or N-1 and N-7 (3a). The observed coordination mode differences is the result of blocking the N-7 and N-9 positions, respectively, by a methyl group. Hence di-, oligo- or polymeric structures (ZnLCl₂)_n can be proposed for 2a and 3a.

For **1a** the shielding of two nitrogens (N-1 and N-7) occurs as well, but its origin must be different. The X-ray data reveal that the purine rings are N-7 bonded to Zn(II). On the other hand, the absence of N-7/N-9 tautomerism, present in **1**, results in the H-9…N-1' hydrogen bonds [20]. Assuming that **1a** exists in dmf- d_7 solution in the same form as in the solid state, one can regard the shielding of N-7 and N-1 as the result of Zn(II) coordination and H-bond formation, respectively.

The observed values of Zn(II)-induced ¹⁵N coordination shifts (3–15 ppm to lower frequency) are similar or slightly larger than those described for Zn(II) chloride complexes with 1,2,4-triazolo- $[1,5\alpha]$ -pyrimidines (max. 2 ppm) [36,37] or adenosine (2–9 ppm) [38]. It is worth noting that the shielding of Zn(II)-coordinated nitrogen atoms is much larger for the imidazole ring (N-7 and N-9, approximately 10–15 ppm) than for the pyrimidine one (N-1, approximately 3–5 ppm).

The values of $J_{\rm NH}$ coupling constants found for **1a** are similar to those of **1**: ${}^{2}J_{\rm N1H2} = 14.5$, ${}^{2}J_{\rm N1H6} = 9.9$, ${}^{2}J_{\rm N3H2} = 15.0$, ${}^{2}J_{\rm N7H8} = 11.2$, ${}^{2}J_{\rm N9H8} = 9.5$, ${}^{4}J_{\rm N3H6} = 1.8$, ${}^{2}J_{\rm N3H8} = 1.7$ Hz.

3.3. ¹⁵N CP MAS spectra of 1 and 1a

In the ¹⁵N CP MAS spectrum of **1** four resonances at -103.3, -115.7, -150.6, -227.6 ppm have been detected (Fig. 4). The two former signals can be assigned





to N-1 and N-3, respectively, by analogy with the spectra measured in dmf- d_7 or dmso- d_6 . The two latter peaks must derive from N-7 or N-9 atoms, the more shielded representing the pyrrole-type nitrogen. Because in the solid state the H-7 tautomer is present [39] (contrary to the solutions, for which the H-9 form is more stable [1]), the signal at -227.6 ppm can be assigned to N-7 and that at -150.6 ppm to N-9. It leads to the following order of ¹⁵N resonances: N-1 > N-3 > N-9 > N-7 (decreasing the chemical shifts), different from that known for **1** in solution.

The ¹⁵N CPMAS spectrum of **1a** exhibits resonances at -115.1, -124.6, -185.8/-188.0, -212.4 ppm (Fig. 4). We can suppose that the signal at -115.1 ppm (close to that at -115.7 ppm in the spectrum of 1) belongs to N-3, as this nitrogen is not engaged in any intra- or intermolecular interactions in both compounds. The peak at -124.6 ppm probably represents N-1, its 21 ppm low-frequency shift being a consequence of the Hbond formation to H-9 [20]. The signal at -212.4 ppm derives from N-9 as in the case of 1a it is a pyrrole-type nitrogen [20]. The twin peaks of smaller but similar intensity at -185.8 and -188.0 ppm must represent the same N-7 atom, coordinated by Zn(II). Thus the suggested order of ¹⁵N resonances for 1a is different from that for 1: N-3 > N-1 > N-7 > N-9 (decreasing the chemical shifts).

The phenomenon of two peaks assigned to N-7 can be explained by a detailed analysis of the X-ray structure of **1a**. Due to the specific packing of molecules in the crystal lattice, the two purine rings in **1a** are not fully equivalent. This effect is especially evident for N-7, for which there are two values of Zn–N bond lengths [Zn–N-7 2.027(3) and Zn–N-7' 2.033(4) Å] and four Cl–Zn–N angles [112.1(1°), 111.7(1°), 111.4(1°), 107.0(1)°] [20]. Hence N-7 and N-7' are located in slightly different moieties which results in the presence of two resonances in the ¹⁵N CP MAS spectrum. When **1a** is dissolved in dmf- d_7 , N-7 and N-7' become chemically and magnetically equivalent, producing one ¹⁵N NMR signal.

4. Conclusions

The Zn(II) chloride complexes with purine, 7-methylpurine or 9-methylpurine differ in stoichiometry and the coordination modes. The reaction of ZnCl₂ with purine in ethanol yields monomeric, N-7-coordinated Zn(purine)₂Cl₂. Analogous syntheses with 7-methylpurine or 9methylpurine produce di-, oligo- or polymeric compounds of the [Zn(methylpurine)Cl₂]_n type.

The Zn(II) coordination of heterocyclic nitrogens results in 3–15 ppm low-frequency shifts of their ¹⁵N NMR signals. This effect indicates N-7; N-1, N-9 and N-1, N-7 nitrogen atoms as the metallation sites in the molecules of purine, 7-methylpurine and 9-methylpur-

ine, respectively. A similar shift of N-1 in $Zn(puri-ne)_2Cl_2$, however, seems to be the result of hydrogen bond formation.

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